Excited state QM/MM dynamics of a BLUF photoreceptor

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Yale

BLUF Photoreceptor

BLUF: blue light using flavin

optogenetics: use light to control cells in living tissue



Source: Getty's Open Content Program (John B. Carnett / Getty Images)



turns on/off

BLUF, coupled to an effector, causes changes under blue light (neuron fires, catalyze a reaction, etc.)

It's a blue light sensitive switch.

BLUF Photoreceptor



BLUF Photoreceptor



SIr1694 BLUF Photocycle

Flavin (FMN) absorbs blue light, form FMN*

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Electron transfer from Tyr to FMN*, form FMN^{•-} (7 ps)

Proton transfer from Tyr to FMN^{•−}, form FMNH[•] (6 ps)

Relaxation to light-adapted state, back to FMN (65 ps)

But, what are changes in the active site?

How can non-equilibrium excited state QM/MM dynamics help understand photocycle?



Gauden, M.; van Stokkum, I. H. M.; Key, J. M.; Lührs, D. C.; van Grondelle, R.; Hegemann, P.; Kennis, J. T. M. PNAS. 2006.

Excited State QM/MM Dynamics

How can excited state QM/MM dynamics help understand photocycle?

Simulate non-equilibrium excited state QM/MM dynamics directly for BLUF SIr1694 starting from locally excited flavin, LE state



Tamm-Dancoff Approximation (TDA) to avoid potential triplet instabilities LRC-ωPBEh chosen after extensive benchmarking vs CASSCF+NEVPT2

Benchmarking DFT vs CASSCF+NEVPT2

Compare PES of double proton transfer with respect to CASSCF+NEVPT2 reference for states of interest (GS, LE, CT)

State	LE			СТ		
Functional	Max Error	Mean Error	RMSD	Max Error	Mean Error	RMSD
LRC- ωPBEh	-0.74	0.16	0.26	-0.57	-0.27	0.31
CAM- B3LYP	0.79	0.18	0.29	-0.96	-0.69	0.71
ωB97X-V	0.97	0.49	0.51	0.93	0.51	0.54
CAM- QTP01	1.03	0.55	0.57	1.08	0.65	0.67

Error in functional with respect to CASSCE+NEVPT2

*includes all grid points, units in eV

- LRC- ω PBEh gave the best agreement
- CAM-B3LYP underestimates CT
- CAM-QTP01 and $\omega B97X\text{-V}$ tend to overestimate both LE and CT states



Sayfutyarova, E. R.; Goings, J. J.; Hammes-Schiffer, S. Electron-Coupled Double Proton Transfer in the SIr1694 BLUF Photoreceptor: A Multireference Electronic Structure Study. *J. Phys. Chem. B* **2019.**

Local heating with QM/MM relaxation¹

QM/MM with excited states can be very sensitive to initial conditions

Zero-point energy (ZPE)

problem: classical MD has no ZPE, but sampling from Wigner distribution impractical for proteins

Shift from classical to QM Hamiltonian

problem: shifting from classical to QM/MM can add additional energy



¹Mai, S.; Gattuso, H.; Monari, A.; González, L. *Front Chem* **2018**, *6*, 495.

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 $= Nk_BT$

 $\implies T = E(\text{ZPE})/Nk_B$

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solution: let system relax for a short period after heating to dissipate energy and move closer to equilibrium bond lengths, etc.

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System preparation



Final QM region temperature: 561 ± 58 K Final MM region temperature: 298 ± 7 K

Figure and schema adapted from Mai, S.; Gattuso, H.; Monari, A.; González, L. Front. Chem. 2018.

1 Excited State QM/MM Dynamics

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100 trajectories, start with population on S1 (locally excited flavin, LE state)



1 Excited State QM/MM Dynamics

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100 trajectories, start with population on S₁ (locally excited flavin, LE state)





Over the course of the dynamics, we follow S₁ adiabatically, but look for trajectories where the **excited state character changes to charge transfer**.

How many trajectories show charge transfer, how long does it take, and what physical factors are involved?







We observe charge transfer for **9 out of 100 trajectories** (within 6 ps). For every trajectory, S_1 is initially of flavin locally excited (LE) character. The character of S_1 changes from LE to CT at the avoided crossing. Neglecting nonadiabatic effects may be one reason for fast CT.

But, what drives charge transfer?







How does protein environment affect CT?



1. Measure electrostatic potential due to environment at Tyr and FMN

$$\Phi(r_i) = \frac{1}{4\pi\epsilon_0} \sum_j \frac{q_j}{r_{ij}}$$

2. Compute potential difference between FMN and Tyr

$$\Delta V = \Phi(r_{\rm FMN}) - \Phi(r_{\rm Tyr})$$

3. More positive $\Delta V(t)$ stabilizes CT



How does protein environment affect CT?



Electrostatic potential increases prior to charge transfer

More positive ΔV \rightarrow lower energy of CT state





What residues influence charge transfer?

Electrostatic potential is additive, so we can decompose into contributions by individual residue; look for residues with greatest variance

Flexible charged groups around the flavin binding pocket: positive and negative (or polar)

Moving net positive charge toward flavin stabilizes CT

But electrostatics are not only governing factor

Energy of charge transfer state depends on the distance between electron donor (Tyr) and acceptor (flavin)

Donor-acceptor distance decreases prior to CT

→ lower energy of CT state

Non-equilibrium response of the protein surrounding the active site can lead to conditions suitable for electron transfer to the flavin

The radical pair Tyr⁺⁺/FMN⁻⁻ drives subsequent proton transfer to neutralize the anionic flavin

Tyr^{•+}/FMN^{•-} radical pair drives proton transfer

Proton transfer from Tyr begins rapidly (20-200 fs) after charge transfer — on the order of a few hydrogen oscillations

Experimentally, tyrosine pK_a changes from 10 to -2 upon oxidation, so first PT is favorable

But, the transfer of protons is correlated with the formation of diradical character in the ground state

Ground state becomes diradical, which requires multireference methods

photocycle progression

TDA-TDDFT no longer suitable!

Spin-flip (SF)-TDA-TDDFT: Designed to treat ground and excited states on equal footing.

Useful in treating radicals, open shell species, and conical intersections.

The ground state shows up as the first "excited state."

So, when reference state fails, pick up where TDA-TDDFT left off but use spin-flip approach — qualitative, but allows us insight into potential mechanism.

Shao, Y.; Head-Gordon, M.; Krylov, A. I. The Spin–flip Approach within Time-Dependent Density Functional Theory: Theory and Applications to Diradicals. *JCP.* **2003**, *118* (11), 4807–4818.

4 Hydrogen Bond Rearrangement

After protonation of flavin, glutamine exists as one of two imidic acid tautomers

Fast glutamine rotation if ZE tautomer, no motion observed for EE (on this timescale)

Recent similar arguments for BLUF domain BlrB: Domratcheva, T.; *et al. Sci. Rep.* **2016**. Grigorenko, B. L.; *et al. Phys. Chem. Chem. Phys.* **2018**.

Summary

Direct simulation of non-equilibrium excited state QM/MM dynamics for SIr1694 BLUF

Electron transfer to flavin driven by protein reorganization and electrostatics

Tyr^{•+}/FMN^{•-} radical pair drives proton transfer

Direct dynamical support for Gln tautomerization + rotation mechanism in SIr1694 photocycle

Acknowledgments

Thank you!

CASSCF+NEVPT2 benchmarking data: Elvira Sayfutyarova

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Extreme Science and Engineering Discovery Environment

Gln50: proton relay favored in Slr1694

- Crystal structure does not provide unambiguous assignment of Gln orientation
- Umbrella sampling used to explore free energy changes along Gln rotation
- Proton relay always favored regardless of Trp_{in} or Trp_{out}

Goings, J. J.; Reinhardt, C. R.; Hammes-Schiffer, S. Propensity for Proton Relay and Electrostatic Impact of Protein Reorganization in SIr1694 BLUF Photoreceptor. *J. Am. Chem. Soc.* **2018.**

Experimental SIr1694 BLUF Photocycle

Suggest changes in hydrogen bonding network around the flavin

Kennis, J. T. M.; Mathes, T. Molecular Eyes: Proteins That Transform Light into Biological Information. Interface Focus 2013.